

## Bespoke Angiogenesis Assay Protocol

### Equipment

37°C 5% CO<sub>2</sub> Incubator  
 Glass Haemocytometer for manual cell counting  
 Light Microscope  
 Laminar Flow Hood  
 Serological pipettes  
 Tissue Culture Flasks

### Materials and Reagents:

#### Cells

[ZHC-5102](#) Human Dermal Fibroblasts (HDFa), Angiogenesis tested, 1 x 10<sup>6</sup> cells/vial.  
[ZHC-2102](#) Human Umbilical Vein Endothelial Cells (HUVEC), Angiogenesis tested, 0.5 x 10<sup>6</sup> cells/vial.

#### Human Fibroblast Growth Medium (FGM) [ZHM-5923](#)

KC1017	Human Fibroblast Basal Medium, 500ml	2-8°C
KC1018	Human Fibroblast Growth Medium Supplement	-20°C

#### Human Large Vessel Endothelial Cell Growth Medium (LVGM) [ZHM-2953](#)

KC1015	Human Large Vessel Endothelial Cell Basal Medium, 500ml	2-8°C
KC1014	Growth Supplement, 10ml	-20°C

#### Angiogenesis Seeding (ASM) and Growth Medium (AGM) [ZHA-1960](#) and [ZHA-1970](#)

KC1012	Angiogenesis Basal Medium, 500ml	2-8°C
KC1013	Angiogenesis Seeding Medium Supplement, 25ml	-20°C
KC1014	Growth Supplement, 10ml	-20°C

#### Angiogenesis Assay Controls [ZHA-1300](#)

KC1007	Suramin Control Compound (1mM)	2-8°C or -20°C
KC1006	VEGF Control Compound (2µg/ml)	2-8°C or -20°C

#### CD31 Staining [ZHA-1225](#)

KC1004	Mouse Anti-Human CD31 Primary Antibody	2-8°C or -20°C
KC1005	Goat Anti-Mouse IgG AP Secondary Antibody	2-8°C or -20°C
KC1010	BCIP/NBT Substrate Tablet	-20°C

Trypsin 1 x solution	SIGMA
Trypsin/EDTA	Sciencell
Buffered saline rinsing solution	Sciencell
Trypsin blocking solution	Sciencell
Trypan Blue	SIGMA

## Method

*Cell culture in the Laminar Flow Hood should be undertaken in strict sterile conditions.*

### Day 1

1. Prepare a bottle of Human Fibroblast Growth Medium by supplementing Human Fibroblast Basal Medium with Human Fibroblast Growth Medium Supplement then equilibrating it to 37°C in a 5% CO<sub>2</sub> incubator for 30 minutes.
2. Prepare a bottle of Human Large Vessel Endothelial Cell Growth Medium by supplementing Human Large Vessel Endothelial Cell Basal Medium with Growth Supplement then equilibrating it to 37°C in a 5% CO<sub>2</sub> incubator for 30 minutes.
3. Thaw the HDFa and HUVEC cells in a water bath.
4. Count the cells under a light microscope using a haemocytometer and the Trypan Blue viability stain.
5. In the laminar flow hood, add the contents of the ampoules into tissue culture flasks containing the relevant medium i.e. FGM for HDFa and LVGM for HUVEC, and place flasks in the 5% CO<sub>2</sub> incubator thereby seeding the cells. Cells should be seeded such that both cell lines are able to be harvested at the same time when less than confluent.

### Day 2

6. After 24 hours, remove the flasks from the incubator and check the cells for growth and sterility under the microscope.
7. In the laminar flow hood, aspirate the medium and replace with fresh FGM and LVGM.
8. Return flasks to the 5% CO<sub>2</sub> incubator.
9. Continue steps 6 to 8 until cells are nearing confluency.

### Day X

10. Prepare a bottle of Angiogenesis Seeding Medium by supplementing Angiogenesis Basal Medium with Angiogenesis Seeding Medium Supplement then equilibrating it to 37°C in a 5% CO<sub>2</sub> incubator for 30 minutes.
11. Before HDFa and HUVEC are confluent, trypsinise the cells according to the V2a Kit protocol.
12. Count cells as in step 4.
13. Plate cells in a 24 well plate into Angiogenesis Seeding Medium at a ratio that promotes suitable endothelial cell tubule formation.
14. Return plate to the 5% CO<sub>2</sub> incubator for 24 hours.

### Day X+1

15. Prepare a bottle of Angiogenesis Growth Medium by supplementing Angiogenesis Basal Medium with Growth Supplement then equilibrating it to 37°C in a 5% CO<sub>2</sub> incubator for 30 minutes.
16. In the laminar flow hood, aspirate the medium and replace with Angiogenesis Growth Medium, containing control (Suramin and VEGF) and test compounds.
17. Return plate to the 5% CO<sub>2</sub> incubator.
18. Continue to change the medium (containing control and test compounds) every two to three days until 14 days have elapsed.

### Day X+14

19. Fix and stain the tubules with the CD31 antibodies according to the V2a Kit protocol in order to analyse.